

Theophylline-sensitive modulation of non-cholinergic excitatory neurotransmission in the guinea-pig ileum

L. Barthó, G. Pethő & Z. Rónai

Department of Pharmacology, University Medical School of Pécs, H-7643 Pécs, Hungary

Atropine-resistant longitudinal contractions of the guinea-pig ileum due to field stimulation were greatly augmented by theophylline (150 μ M). Adenosine exerted an opposite effect. Theophylline did not potentiate contractions evoked by exogenous substance P. It is suggested that a theophylline-sensitive inhibitory mechanism (possibly mediated by adenosine or a related substance) controls transmitter release from enteric non-cholinergic excitatory neurones.

Introduction Theophylline increases the stimulation-evoked release of neurotransmitters in many experimental situations; among others it facilitates the release of acetylcholine in the myenteric plexus (for review see Vizi, 1979; Fredholm, 1980). The aim of the present study was to test the effect of theophylline on atropine-resistant longitudinal contractions of the field-stimulated, isolated ileum of the guinea-pig (Ambache & Freeman, 1968). These responses are most probably mediated by a substance P-like peptide, released from intrinsic neurones of the myenteric plexus (cf. Furness & Costa, 1980; Barthó & Holzer, 1985). The effect of adenosine, the most likely endogenous activator of theophylline-sensitive (P_1) purinoceptors (Burnstock, 1978), was also investigated.

Methods Segments of guinea-pig ileum of approximately 2 cm length were suspended in Krebs-Henseleit solution of the following composition (mM): NaCl 118.3, KCl 4.7, $CaCl_2$ 2.5, $MgSO_4$ 1.2, $NaHCO_3$ 25, KH_2PO_4 1.2 and glucose 10. The solution was kept at 37°C and aerated with 5% CO_2 and 95% O_2 . Longitudinal contractions were recorded with the aid of an isotonic transducer (low-friction tuning capacitor system; signal: 1–100 mV) connected to a compensographic recorder (MTA-KUTESZ, Hungary). The load on the tissue was 0.005 N (0.5 g). Naloxone (0.5 μ M) and guanethidine (5 μ M) were also added to the organ bath to prevent opioid and sympathetic inhibition of non-cholinergic excitatory neurotransmission (cf. Barthó & Holzer, 1985). Field stimulation was applied through a pair of longitudinal platinum electrodes. Parameters of stimulation were:

supramaximal voltage, 0.1 ms impulse width; 150 impulses were delivered at a frequency of 1, 5 or 20 Hz. In each experiment only one of these frequencies was used and stimulations were separated by 10-min intervals. Drug effects were investigated only when uniform control responses had been obtained. Contact time for adenosine (Reanal), theophylline (Sigma) and tetrodotoxin (Sankyo) was 3 min. In some experiments strips of myenteric plexus-longitudinal muscle were used; they were prepared according to the method of Paton & Vizi (1969). Contractions were expressed as % of the maximal response evoked by histamine (5 μ M) given at the end of the experiment. Data are expressed as mean \pm s.e.mean. Student's one-sample *t* test was used for assessing the statistical significance of differences.

Results Atropine-resistant ileum contractions evoked by field stimulation were potently enhanced by theophylline (150 μ M). Smaller concentrations of theophylline (15 and 50 μ M) had similar effects; however, an increase in responses to field stimulation was still evident when the concentration of theophylline was raised from 50 to 150 μ M ($n = 5$). Half-maximal contractions produced by substance P (1–3 nM) were slightly but significantly reduced by theophylline (150 μ M; Figure 1). Adenosine inhibited non-cholinergic contractions; this effect was prevented by theophylline (Figure 1) in a surmountable fashion. Theophylline (150 μ M) caused an apparently parallel shift to the right of the concentration-response curves for adenosine. Estimated EC_{50} values of adenosine on contractions due to 5 Hz stimulation were 6.9 ± 0.8 μ M in the absence and 46.8 ± 4.0 μ M in the presence of theophylline (150 μ M; $n = 9$). The effects of both theophylline and adenosine were readily reversible by washing. Adenosine (up to 50 μ M) did not inhibit the action of exogenous substance P which was used in concentrations (1–3 nM) producing approximately half-maximal contractions. In the presence of adenosine (50 μ M) these contractions reached $99 \pm 3.7\%$ of those in the absence of adenosine ($n = 10$). Theophylline (150 μ M) also enhanced and adenosine (15 μ M) also inhibited non-cholinergic contractions due to field stimulation (5 Hz) of myenteric plexus-

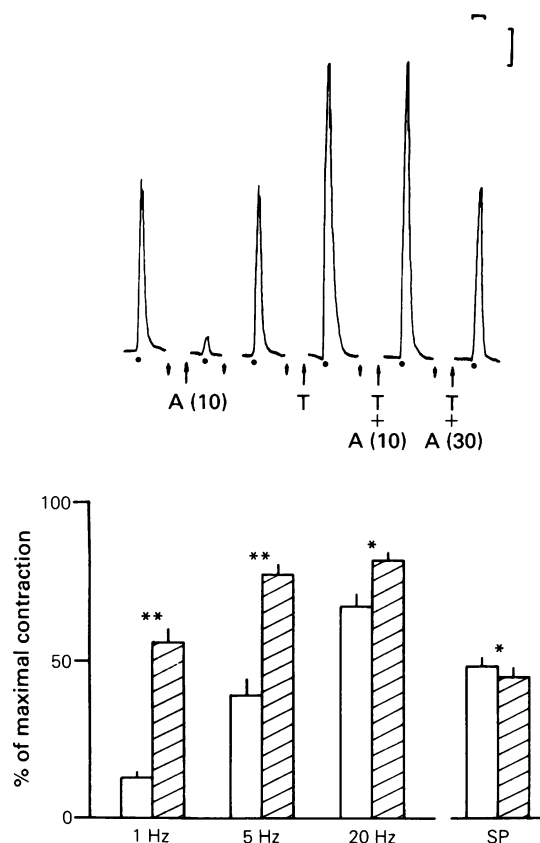


Figure 1 (a) Atropine-resistant longitudinal contractions of the guinea-pig ileum due to field stimulation (dots; 150 impulses at 5 Hz). The effect of adenosine (A) in the absence and in the presence of theophylline (T; 150 μ M). Concentrations of adenosine (μ M) are indicated by the numbers in parentheses. At downward arrows the bath fluid was changed three times. Calibrations: 10% of the maximal contraction produced by 5 μ M of histamine (vertical); 1 min (horizontal). (b) Effect of theophylline on atropine-resistant contractions due to field stimulation at different frequencies ($n = 8$ in each group) or in response to substance P (SP; 1–3 nM, $n = 10$). Open columns: control response. Hatched columns: responses in the presence of theophylline (150 μ M). Mean values are given with s.e. mean shown by vertical lines. Significant changes are indicated by asterisks; * $P < 0.05$, ** $P < 0.001$.

longitudinal muscle preparations ($n = 5$). On segments of whole ileum, half-maximally contracted by histamine, transmural stimulation (single pulses at 0.1 Hz or 5 pulses at 5 Hz) produced rapid and short-lived relaxations. These responses were not influenced by theophylline (150 μ M, $n = 5$). Tetrodotoxin (0.5 μ M) prevented all of the above responses to field stimulation ($n = 4$ each).

Discussion These findings provide pharmacological evidence for the presence of a theophylline-sensitive modulation of transmitter release from non-cholinergic excitatory neurones of the myenteric plexus. The stimulant action of theophylline on the atropine resistant contractile responses is unlikely to be explained by a post-junctional potentiation of smooth muscle contraction or by the elimination of a nerve-mediated non-adrenergic relaxation mechanism since contractions due to exogenous substance P were not enhanced and non-adrenergic relaxations due to field stimulation were not inhibited by theophylline. Likewise, in the guinea-pig taenia caeci, theophylline does not inhibit the non-adrenergic relaxation due to field stimulation (Small & Weston, 1979). We suggest that theophylline enhances contractions of the ileum to field stimulation by eliminating the action of an endogenous inhibitory substance and thus enhancing the release of the non-cholinergic excitatory mediator. Our findings with adenosine, in conjunction with those of others (Burnstock, 1978; Vizi, 1979) may indicate that the inhibitory substance is adenosine or another adenine derivative. EC_{50} values for adenosine (in the absence and in the presence of theophylline) are similar to those reported by Sawynok & Jhamandas (1976) on cholinergic 'twitch' responses of the guinea-pig ileum. The present results support the view (Barthó & Holzer, 1985) that the pharmacology of the enteric release of acetylcholine and substance P is similar.

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